

DEVELOPMENT OF A FUNCTIONALIZED MCM-41 BIOSENSOR MODIFIED
WITH SELENIUM NANOPARTICLES FOR GLUCOSE DETERMINATION

NURUL ASYIKIN BINTI KAMARUZAMAN

A thesis submitted in fulfillment of the
requirements for the award of the degree of
Master of Science (Chemistry)

Faculty of Science
Universiti Teknologi Malaysia

OCTOBER 2012

DEVELOPMENT OF A FUNCTIONALIZED MCM-41 BIOSENSOR MODIFIED
WITH SELENIUM NANOPARTICLES FOR GLUCOSE DETERMINATION

NURUL ASYIKIN BINTI KAMARUZAMAN

A thesis submitted in fulfillment of the
requirements for the award of the degree of
Master of Science (Chemistry)

Faculty of Science
Universiti Teknologi Malaysia

OCTOBER 2012

Special dedications to

My beloved:

Dad and Mum..... Kamaruzaman Abd Rasid and Nor Arbaayah Sharif

Brothers..... Nor Zaidy, Nor Zaid, Nor Aizat and Nor Hazmi

Sisters in Law.... Sabrina Bt. Sabri and Noor Shahidah Satar

My respected project supervisor

Prof. Alias Mohd Yusof

Co-Supervisors

Prof. Noor Aini Abdul Rashid

Assoc. Prof. Dr. Abdull Rahim Mohd Yusoff

And my dearest friends.....

Thank you for always helping and supporting me

ACKNOWLEDGEMENTS

First and foremost, Alhamdulillah, I am grateful to Allah S.W.T for giving me strength, confidence and patience to overcome problems and obstacles during the course of my study. I would like to express my highest gratitude and appreciation to my main supervisor, Prof. Alias Mohd Yusof for his support, guidance, encouragement and tolerance. To my co-supervisors, Prof. Noor Aini Abdul Rashid for the extensive editing and discussion and Assoc. Prof. Dr. Abdull Rahim Mohd Yusoff for the voltammetric study. Your motivation, advice, patience, and valuable suggestions are really appreciated. Thanks are also due to Dr. Shafinaz Shahir for the input and suggestions on work related to enzymes.

I am also very thankful to members of my research team especially, Kogubalan Illansolan, Mohd Lokman Ibrahim, Shakil Mohammad Arif, Yuhanness Mohd Yusof and Mohd Dzulkhakim Wirzal for giving me the motivation, sharing their knowledge, providing me assistance and the regular discussions which really helped me in my lab work.

Thanks to all my close friends and Radiochemistry lab mates who had accompanied me and encouraged me along this journey. Without their help and support, it would have been a very difficult journey. I would also like to express my appreciation to all laboratory staff from the Department of Chemistry, Faculty of Science especially to Mr Rahim and Mr Hanan for their assistance.

Most of all, my sincere appreciation to my beloved family, whose patience, undying support and everlasting love had kept me motivated. Not forgetting my supportive and caring best friend, Mr Hafsham Mohd Ali for the motivation, support, encouragement, advice and understanding. Last but not least, praise be to Allah Almighty for His inspiration and guidance.

PREFACE

In this work, several reports have been published and presented in the conferences and journals, as listed below:

1. Alias Mohd Yusof, Abdull Rahim Mohd Yusoff, Noor Aini Abdul Rashid, Shafinaz Shahir and Nurul Asyikin Kamaruzaman. *Effect of Temperature and Calcination Time in Characterizations of MCM-41*. Second International Conference and Workshops on Basic and Applied Sciences and Regional Annual Fundamental Science Seminar 2009 (ICORAFSS 2009), The ZON Regency Hotel, Johor Bahru, Johor, 2-4 June 2009.
2. Alias Mohd Yusof, Abdull Rahim Mohd Yusoff, Noor Aini Abdul Rashid, Shafinaz Shahir and Nurul Asyikin Kamaruzaman. *Synthesis of selenium particles in the presence of MCM-41 at different aging temperature*. 10th Asian Conference on Analytical Sciences 2009 (ASIANALYSIS X). PWTC, Kuala Lumpur, 11th – 13th August 2009.
3. Alias Mohd Yusof, Abdull Rahim Mohd Yusoff, Noor Aini Abdul Rashid, Shafinaz Shahir and Nurul Asyikin Kamaruzaman. *Synthesis and Characterization of Selenium Nanoparticles Induced by Ultrasonication Irridiation with Ascorbic Acid as Reducing Agents*. Faculty Science Post-Graduate Conference (FSPGS) 2010, UTM Skudai, Johor, 2010.
4. Alias Mohd Yusof, Abdull Rahim Mohd Yusoff, Noor Aini Abdul Rashid, Shafinaz Shahir and Nurul Asyikin Kamaruzaman. *Glucose Oxidase-Functionalized-Selenium Nanoparticles-Mesoporous Silica as a Glucose Biosensor (HyperSiliSel)*. 12th Industrial Art and Technology Exhibition 2010

(INATEX 2010). 5-7 August 2010, Dewan Sultan Iskandar, Universiti Teknologi Malaysia.

5. Alias Mohd Yusof, Abdull Rahim Mohd Yusoff, , Shafinaz Shahir, Shakil Mohammad Arif, Nurul Asyikin Kamaruzaman and Nik Ahmad Nizam Nik Malek. *A Cholesterol Biosensor Based on Cholesterol Oxidase Immobilized onto Functionalized Mesoporous Silica (CHOL-E-SENSE)*. 12th Industrial Art and Technology Exhibition 2010 (INATEX 2010). 5-7 August 2010, Dewan Sultan Iskandar, Universiti Teknologi Malaysia.
6. Silver Award in 12th Industrial Art and Technology Exhibition 2010 (INATEX 2010). 5-7 August 2010, Dewan Sultan Iskandar, Universiti Teknologi Malaysia for research project of *A Cholesterol Biosensor Based on Cholesterol Oxidase Immobilized onto Functionalized Mesoporous Silica (CHOL-E-SENSE)*.

ABSTRACT

A novel biosensor for glucose determination had been developed in this study. Glucose biosensor is a good example of a commercial biosensor. It uses glucose oxidase (GO_x), a redox enzyme to break down glucose to hydrogen peroxide and coupled with amperometric detection. For the construction of such a biosensor, a broad applicable method in the immobilization of enzyme is critically needed. One way to go about this is through the development of a new carrier such as nanoparticles. Here, a silica-based material, MCM-41, was used as enzyme support material, functionalized and modified with selenium (Se) nanoparticles and then fabricated into a biosensor. MCM-41 was synthesized and characterized to analyze the structural, morphological, elemental and physicochemical characteristics. It was later confirmed that MCM-41 of high purity and high surface area was synthesized. Pristine and unmodified MCM-41 may not be suitable as enzyme support material because it cannot provide the necessary sites for enzyme attachment. Therefore, two types of MCM-41 supports were produced: *f*-MCM-41 and *f*-SeMCM-41. The first one, *f*-MCM-41 was modified for immobilization of GO_x and minimum leaching of the enzyme by functionalizing with amino groups using 3-aminopropyl triethoxysilane (APTES), followed by attachment of aldehyde group using glutaraldehyde. The latter, *f*-SeMCM-41, was co-functionalized with amino group during selenium nanoparticles (SNs) attachment onto the silicate framework to increase sensitivity and electrical conductivity for a better response. The product was then functionalized with glutaraldehyde. Selenium nanoparticles (SNs) were successfully synthesized using a simple, cost effective and non-hazardous procedure where selenious acid was reduced using ascorbic acid, ultrasonicated and aged for 24h. Characterization showed that SNs of hexagonal crystalline type with high purity of more than 95.0% was produced. The incorporation process of SNs onto MCM-41 did not alter the structure of MCM-41 or even the SNs as observed by X-Ray Diffraction Spectroscopy (XRD). It was found that GO_x -*f*-Se-MCM-41 was more efficient than GO_x -*f*-MCM-41 as determined by the specific activity of GO_x immobilized onto them. The optimum pH for immobilization of GO_x onto both functionalized MCM-41 and Se-MCM-41 was determined to be pH 6.0 and the optimum initial GO_x concentration was 2.0 mg/mL. GO_x -*f*-MCM-41 and GO_x -*f*-Se-MCM-41 were used in the fabrication of carbon paste electrodes (CPE) and the efficiency examined. GO_x -*f*-Se-MCM-41/CPE electrode was more sensitive and efficient as compared to GO_x -*f*-MCM-41/CPE electrode, as evaluated using cyclic voltammetry. GO_x -*f*-Se-MCM-41/CPE can detect very low range of glucose between 3.69 μM to 16.25 μM . Normal human glucose level is between 3.3 to 3.8 mM but this biosensor can detect much lower levels making it an excellent biosensor for clinical and industrial use. Hence, the newly developed functionalized MCM-41 support with immobilized glucose oxidase with Se attached to it, GO_x -*f*-Se-MCM-41/CPE offers the potential exploitation of a suitable glucose biosensor.

ABSTRAK

Suatu biosensor baru telah dibangun dalam kajian ini bagi penentuan glukosa. Biosensor glukosa adalah contoh yang terbaik untuk biosensor komersial. Ia menggunakan oksidasa glukosa (GO_x), enzim redoks bagi glukosa untuk memecahkannya kepada hidrogen peroksida dan digabungkan dengan pengesanan amperometrik. Satu kaedah yang mudah bagi menyahgerakkan enzim amat diperlukan untuk pembinaan biosensor. Salah satu cara adalah melalui pembangunan pengangkut baru seperti nanopartikel. Di sini, bahan berasaskan silika, MCM-41 telah digunakan sebagai bahan sokongan enzim, difungsikan dan diubahsuai dengan selenium (Se) nanopartikel (SNs) yang kemudiannya direka menjadi biosensor. MCM-41 telah disintesis dan dicirikan untuk menganalisis struktur, morfologi, unsur dan ciri-ciri fizikokimia. MCM-41 yang telah disintesis disahkan mempunyai ketulenan yang tinggi dan luas permukaan yang besar. MCM-41 yang asli dan tidak diubahsuai tidak sesuai sebagai bahan sokongan enzim kerana ianya tidak dapat menyediakan tapak yang diperlukan untuk sokongan enzim tersebut. Oleh itu, dua jenis MCM-41 telah dihasilkan: *f*-MCM-41 dan *f*-Se-MCM-41. Yang pertama, *f*-MCM-41 telah diubahsuai bagi pergerakan GO_x menunjukkan larut lesap enzim yang minimum apabila difungsikan dengan kumpulan amino menggunakan 3-aminopropil trietoksilana (APTES), diikuti oleh pelekatan kumpulan aldehid menggunakan glutaraldehid. Kedua, *f*-SeMCM-41, difungsikan seperti di atas tetapi dengan pengubahsuaian selanjutnya melalui penggabungan SNs ke dalam rangka kerja silikat untuk meningkatkan kepekaan dan kekonduksian elektrik bagi tindak balas yang lebih baik. Kemudian, produk telah difungsikan dengan glutaraldehid. SNs telah berjaya disintesis dengan menggunakan kaedah yang mudah, kos yang efektif dan tidak berbahaya di mana asid selenious telah diturunkan dengan menggunakan asid askorbik, diultrasonik dan disimpan di tempat gelap selama 24 jam. Pencirian menunjukkan bahawa SNs jenis kristal heksagon dengan ketulenan yang tinggi melebihi 95.0% telah dihasilkan. Proses penggabungan SNs ke MCM-41 tidak mengubah struktur MCM-41 atau SNs seperti yang ditunjukkan oleh Pembelauan sinar-X (XRD). Ia mendapati bahawa GO_x -*f*-Se-MCM-41 adalah lebih berkesan daripada GO_x -*f*-MCM-41 seperti yang dihitung oleh aktiviti spesifik GO_x yang dinyahgerak ke atas bahan sokongan ini. pH optimum untuk pergerakan GO_x ke atas kedua-dua MCM-41 atau Se-MCM-41 difungsi telah ditetapkan pada pH 6.0 dan kepekatan awal GO_x optimum adalah 2.0 mg/mL. GO_x -*f*-MCM-41 dan GO_x -*f*-Se-MCM-41 telah digunakan dalam fabrikasi elektrod pasta karbon (CPE) dan kecekapannya dikaji. Elektrod GO_x -*f*-Se-MCM-41/CPE adalah lebih peka dan berkesan berbanding GO_x -*f*-MCM-41/CPE elektrod, seperti yang dianalisis dengan menggunakan voltammetri kitaran. GO_x -*f*-Se-MCM-41/CPE boleh mengesan glukosa yang sangat rendah antara 3.69 μM ke 16.25 μM . Paras glukosa manusia normal adalah di antara 3.3-3.8 mM tetapi biosensor ini dapat mengesan pada tahap yang lebih rendah menjadikannya biosensor yang sangat baik untuk kegunaan klinikal dan industri. Oleh itu, MCM-41 sokongan yang baru dibangun telah difungsikan dengan GO_x dinyahgerakkan dengan Se terlekat padanya, GO_x -*f*-Se-MCM-41/CPE, berpotensi untuk dieksplotasi sebagai biosensor glukosa.

TABLE OF CONTENTS

CHAPTER	TITLE	PAGE
	DECLARATION	ii
	DEDICATION	iii
	ACKNOWLEDGEMENTS	iv
	PREFACE	v
	ABSTRACT	vii
	ABSTRAK	viii
	TABLE OF CONTENTS	ix
	LIST OF TABLES	xvi
	LIST OF FIGURES	xvii
	LIST OF ABBREVIATIONS	xxi
	LIST OF APPENDICES	xxiii
1	Introduction	1
	1.1 Background of Study	1
	1.2 Problem Statements	4
	1.3 Objectives of Study	5
	1.4 Scope of Study	5
	1.5 Significance of Study	6

2	Literature Review	7
2.1	Diabetes Mellitus	7
2.2	Mesoporous Materials	8
2.2.1	MCM-41	9
2.2.2	Modification of MCM-41 with Metals	10
2.3	Selenium	12
2.3.1	Allotropes of Selenium	14
2.3.1.1	Vitreous Selenium	14
2.3.1.2	α - and β -monoclinic Selenium	14
2.3.1.3	Trigonal Selenium	15
2.3.2	Selenium Nanoparticles	16
2.4	Enzyme Immobilization	16
2.4.1	Adsorption	21
2.4.2	Entrapment	22
2.4.3	Covalent Binding	23
2.4.4	Cross-linking	23
2.4.5	Mesoporous Silica as Support for Enzyme Immobilization	25
2.5	Electrochemical Sensors for Clinical Analysis	28
2.5.1	Glucose Biosensors	29
3	Materials and Methods	33
3.1	Chemicals and Reagents	33

3.2	Instruments	34
3.3	Synthesis of MCM-41	35
3.4	Synthesis of Selenium Nanoparticles (SNs)	35
3.5	Incorporations of SNs with MCM-41 in the Presence of APTES.	36
3.6	Leaching Studies	36
3.6.1	Decomposition of Se-MCM-41	37
3.6.2	Leaching Study of Se-MCM-41	38
3.7	Functionalization of MCM-41 with Amino Group	38
3.8	Functionalization of MCM-41-A and Se-MCM-41 with Aldehyde Group.	39
3.8.1	Tollen's Reagent Method.	39
3.9	Characterizations of the MCM-41, SNs and Se-MCM-41.	40
3.9.1	X-Ray Diffraction Spectroscopy Analysis	40
3.9.2	FTIR Spectroscopy	41
3.9.3	Field Emission Scanning Electron Microscope	41
3.9.4	Thermal Electron Microscope	42
3.9.5	Thermal Gravimetric Analysis and Differential Thermal Analysis	42
3.10	Optimization of Immobilization of Glucose Oxidase onto <i>f</i> -MCM-41 and <i>f</i> -Se-MCM-41	42
3.10.1	Effect of pH for Immobilization of Glucose Oxidase onto <i>f</i> -MCM-41 and <i>f</i> -Se-MCM-41.	43
3.10.2	Effect of GO _x Concentration Used for the Immobilization of Glucose Oxidase onto <i>f</i> -MCM-41 and <i>f</i> -Se-MCM-41.	43

3.11	Preparation of Reagents for the Determination of Enzymatic Assay	44
3.11.1	Preparation of Sodium Acetate Buffer (Reagent A)	44
3.11.2	Preparation of o-Dianisidine Solution (Reagent B)	44
3.11.3	Preparation of β -D (+) Glucose Substrate Solution (Reagent C)	45
3.11.4	Preparation of Reaction Cocktail (Reagent D)	45
3.11.5	Preparation of Peroxidase Enzyme Solution (Reagent E)	45
3.12	Determination of Enzymatic Assay using Continuous Spectrophotometric Rate Determination	45
3.13	Determination of Protein Concentration using Bradford Method	47
3.13.1	Preparation of Standard 3.1 mL Assay Protocol	47
3.14	Determination of Specific Activity of Glucose Oxidase	47
3.15	Leaching Test of the Immobilized <i>f</i> -MCM-41 and <i>f</i> -Se-MCM-41	48
3.16	Immobilization of GO_x onto <i>f</i> -MCM-41 and <i>f</i> -Se-MCM-41.	48
3.17	Preparation of Working Electrode	49
3.18	Study on Electrochemical Properties of Glucose Oxidase-Selenium Nanoparticles-MCM-41.	49
3.18.1	Effect of pH	50
3.18.2	Effect of Scan Rate	50
3.19	Sample Analysis Using Glucose Biosensor	50

4	Synthesis and Characterization of MCM-41, Se-MCM-41 and Glutaraldehyde Functionalized Se-MCM-41	51
4.1	Introduction	51
4.2	Synthesis and Calcination of MCM-41	52
4.3	Characterization of MCM-41	53
4.3.1	Analysis of MCM-41 with X-Ray Diffraction Spectroscopy	53
4.3.2	Analysis of MCM-41 with Fourier Transform Infrared Spectroscopy.	56
4.3.3	Analysis of MCM-41 with Thermal Gravimetric Analysis	58
4.3.4	Analysis of MCM-41 with Field Emission Scanning Electron Microscopy	59
4.3.5	Analysis of MCM-41 with Thermal Emission Microscopy	60
4.4	Synthesis of Se-nanoparticles (SNs)	60
4.5	Characterization of SNs	63
4.5.1	Analysis of SNs with X-Ray Diffraction Spectroscopy	63
4.5.2	Analysis of SNs with Thermal Gravimetric Analysis	64
4.5.3	Analysis of SNs with Field Emission Scanning Electron Microscopy	65
4.6	Incorporation of SNs onto MCM-41	66
4.7	Characterization of Se-MCM-41	67
4.7.1	Analysis of Se-MCM-41 with X-Ray Diffraction Spectroscopy.	68
4.7.2	Analysis of Se-MCM-41 with Fourier Transform Infrared.	68

4.7.3	Analysis of Se-MCM-41 with Thermal Gravimetric Analysis	69
4.7.4	Analysis of Se-MCM-41 with Field Emission Scanning Electron Microscopy	71
4.8	Leaching Study of Se-MCM-41	72
4.9	Functionalization of MCM-41 with Amino and Aldehyde Group and its Characterization	73
4.9.1	Analysis of MCM-41, MCM-A and <i>f</i> -MCM-41 with XRD Spectroscopy	75
4.9.2	Analysis of MCM-41, MCM-A and <i>f</i> -MCM-41 with FTIR	76
4.10	Functionalization of Se-MCM-41 with Aldehyde Group and its Characterization	78
4.10.1	Analysis of <i>f</i> -Se-MCM-41 using XRD Spectroscopy	78
4.10.2	Analysis of <i>f</i> -Se-MCM-41 using FTIR Spectroscopy	80
5	Optimization of Glucose Oxidase Immobilization onto MCM-41 Support	82
5.1	Immobilization of Glucose Oxidase (GO _x) onto Functionalized MCM-41 and Se-MCM-41.	82
5.2	Optimization of GO _x Immobilization	83
5.2.1	Effect of pH onto GO _x Immobilization	83
5.2.2	Effect of Initial Concentration of GO _x enzymes	90
5.3	Immobilization of GO _x onto <i>f</i> -MCM-41 and <i>f</i> -Se-MCM-41.	93
5.4	Rationale Behind Why GO _x - <i>f</i> -Se-MCM-41 is Better Than GO _x - <i>f</i> -MCM-41	95

6	Development of Biosensor for Voltammetric Analysis of Glucose	98
6.1	Electrochemical Behaviour of the $\text{GO}_x\text{-f-MCM-41}$ and $\text{GO}_x\text{-f-Se-MCM-41}$.	98
6.1.1	Effect of pH onto $\text{GO}_x\text{-f-MCM-41/CPE}$	99
6.1.2	Effect of Scan Rate onto $\text{GO}_x\text{-f-MCM-41/CPE}$	106
6.1.3	Analysis of $\text{GO}_x\text{-f-MCM-41}$ and $\text{GO}_x\text{-f-Se-MCM-41}$ Carbon Paste Electrodes at Optimum Conditions	109
6.1.4	Analysis of $\text{GO}_x\text{-f-Se-MCM-41/CPE}$ Under Reduced Oxygen Condition	113
7	Conclusions	120
7.1	Conclusions	120
7.2	Suggestions and Recommendations	123
	REFERENCES	126
	APPENDIX	147

LIST OF TABLES

TABLE NO.	TITLE	PAGE
2.1	Some of the methods reported for the formation of different types of Se nanoparticles.	17
2.2	Comparison of performance factors of some commercially available glucose biosensors	30
4.1	Infrared data interpretation for the as-synthesized and calcined MCM-41.	57
4.2	EDX analyses for MCM-41, SNs and Se-MCM-41	72
4.3	FTIR data interpretation for MCM-41-C, MCM-41-A and <i>f</i> -MCM-41	77
4.4	Infrared data interpretation for the comparison of Se-MCM-41 and <i>f</i> -Se-MCM-41	81
6.1	Comparison of response for voltammetry detection at pH 6.0 and 7.0.	104
6.2	Effect of scan rate using DPSV voltammetry	108
6.3	Comparison of GO_x - <i>f</i> -MCM-41/CPE and GO_x - <i>f</i> -Se-MCM-41/CPE using cyclic voltammetry at pH 6.0 with scan rate 10 mV/s.	112
6.4	GO_x - <i>f</i> -Se-MCM-41/CPE with the reduced O_2 in graphite as working electrode at pH 6.0 with scan rate 10 mV/s using linear voltammetry.	116
6.5	Comparison of GO_x - <i>f</i> -Se-MCM-41/CPE with other electrodes	118

LIST OF FIGURES

FIGURE NO.	TITLE	PAGE
2.1	Different types of MCM based on the shape	10
2.2	Structure of monoclinic Se	15
2.3	Structure of trigonal Se	15
2.4	Simple adsorption of enzymes onto surface of support materials	22
2.5	Entrapment of enzymes in gel, film or membrane	22
2.6	Direct covalent bonding between enzyme and support material	23
2.7	Cross-linking of enzyme onto support materials	25
2.8	Clinical analysis procedures based on electrochemical sensors	28
2.9	The principle schematic of (a) first, (b) second and (c) third generation biosensors	32
4.1	Flow chart of procedures involved in the functionalization of MCM-41 and it's incorporation with SNs	52
4.2	XRD pattern of (a) MCM-41-as and MCM-41-C for (b) 1 day (c) 2 days and (d) 3 days of calcination time.	54
4.3	Comparison of slow and rapid ramping time during the calcination process.	55
4.4	Schematic mechanism pathway of the formation of MCM-41	56
4.5	Infrared spectra of (a) as-synthesized and (b) calcined MCM-41	56
4.6	TGA-DTG thermogram of MCM-41	58

4.7	Images of FESEM of MCM-41 at (a) 10 000 (b) 15 000 and (c) 25 000 times magnification	59
4.8	The TEM images of MCM-41 at (a) 25 000 and (b) 100 000 times magnification	60
4.9	Changes in solution colour during transformation process of SNs (a) after addition of ascorbic acid and after (b) 1 h (c) 2 h (d) and 3 h of ultrasonication	61
4.10	XRD pattern of SNs	63
4.11	TGA-DTG thermograms of SNs samples	64
4.12	Comparison FESEM image of (a) <i>a</i> -Se and formation of SNs (b) 24 h aging, (c) using ultrasonication without aging and (d) using ultrasonication and aging in combination	66
4.13	Comparison of XRD patterns in the range of 1.5-70° for (a) Se-MCM-41, (b) SNs and (c) MCM-41	68
4.14	Infrared spectra of (a)MCM-41, (b) SNs and (c) Se- MCM-41	69
4.15	TGA thermograms showing comparison of (a) SNs, (b) MCM-41 and (c) Se-MCM-41	70
4.16	FESEM images of (a) MCM-41 (b) SNs and (c) Se- MCM-41	71
4.17	Plot of amount of Se released from Se-MCM-41 <i>vs</i> leaching period over 24 h	73
4.18	Reaction of MCM-41 with APTES to give MCM-41-A	74
4.19	Reactions involving MCM-41-A with glutaraldehyde to form <i>f</i> -MCM-41	74
4.20	XRD patterns of (a) MCM-41, (b) MCM-41-A and (c) <i>f</i> -MCM-4	75
4.21	FTIR spectra of (a) MCM-41(b) MCM-41-A and (c) <i>f</i> - MCM-41 in the range of 4000-450 cm ⁻¹	76
4.22	Comparison of XRD patterns of (a) Se-MCM-41 and (b) <i>f</i> -Se-MCM-41 in the range 1.5-10°	79
4.23	Comparison of XRD pattern of (a) Se-MCM-41 and (b) <i>f</i> -Se-MCM-41 in the range 10-70°	79
4.24	Comparison of XRD patterns of (a) Se-MCM-41 and (b) <i>f</i> -Se-MCM-41 in the range 1.5-70°	79

4.25	FTIR spectra of (a) Se-MCM-41 and (b) <i>f</i> -Se-MCM-41 in the range of 4000-450 cm ⁻¹	80
5.1	Effects of pH on enzyme specific activity for free, immobilized enzymes and GO _x leaching from the immobilized supports	84
5.2	Schematic diagram of substrate attachment to the enzyme active sites of enzyme immobilized onto MCM-41. Not all the active sites as shown in red circle can be accessed due to the undesired configuration by which the enzyme was positioned	86
5.3	Effect of pH on the percentage specific activity of immobilized enzymes and leaching from the immobilized supports	88
5.4	Effects of initial concentration on GO _x on enzyme specific activity of free, immobilized enzymes and leaching from the immobilized supports	90
5.5	Effect of initial concentration on the percentage specific activity of immobilized enzymes and leaching from the immobilized supports	92
5.6	Amount of GO _x bound onto <i>f</i> -MCM-41 and <i>f</i> -Se-MCM-41 at pH 6.0 with 2.0 mg/mL of GO _x	94
5.7	Percentage of GO _x bound onto <i>f</i> -MCM-41 and <i>f</i> -Se-MCM-41 at pH 6.0 with 2.0 mg/mL of initial concentration of GO _x	94
5.8	Hypothetical diagram showing GO _x binding with the aldehyde groups as well as on SNs (red circle) during GO _x immobilization. The red circle at the far right shows GO _x sandwiched in between SN and aldehyde group which prevents leaching of GO _x	96
6.1	DPCSV of effect of pH in the absence of glucose; $t_{acc}=30$ s, $E_{acc}=+0.2$ V and $E_i=+0.7$ V. Scan rate = 50 mV/s	100
6.2	DPCSV of comparison in the absence (a,c,e,g) and presence (b,d,f,h) of glucose based on pH; $t_{acc}=30$ s, $E_{acc}=+0.2$ V and $E_i=+700$ mV. Scan rate = 50 mV/s	101
6.3	DPCSV voltammograms of glucose at pH 6.0; $t_{acc}=30$ s, $E_{acc}=+0.2$ V and $E_i=+0.7$ V. Scan rate = 50 mV/s	102
6.4	DPSV voltammograms of glucose at pH 7.0; $t_{acc}=30$ s, $E_{acc}=+0.2$ V and $E_i=+0.7$ V. Scan rate = 50 mV/s	102

6.5	Linear fit of DPCSV responding current and concentration of glucose at pH 6.0 and 7.0	103
6.6	DPCSV voltammograms of glucose at pH 5.0; $t_{acc}=30$ s, $E_{acc}=+0.2$ V and $E_i=+0.9$ V. Scan rate = 50 mV/s	104
6.7	DPCSV voltammograms of glucose at pH 9.0. ; $t_{acc}=30$ s, $E_{acc}=+0.2$ V and $E_i=+0.9$ V. Scan rate = 50 mV/s	104
6.8	Linear fit of DPCSV responding current and concentration of glucose working at pH 5.0 and 9.0.	105
6.9	DPCSV voltammograms of glucose with scan rate 10 mV/s; $t_{acc}=30$ s, $E_{acc}=+0.2$ V and $E_i=+0.7$ V	106
6.10	DPCSV voltammograms of glucose with scan rate 30 mV/s; $t_{acc}=30$ s, $E_{acc}=+0.2$ V and $E_i=+0.7$ V	107
6.11	DPCSV voltammograms of glucose with scan rate 50 mV/s; $t_{acc}=30$ s, $E_{acc}=+0.2$ V and $E_i=+0.7$ V	107
6.12	Linear fit of DPCSV responding current and concentration of glucose at scan rate affected by scan rate	108
6.13	Cyclic voltammogram GO_x-f -MCM 41/CPE at pH 6.0 with scan rate 10 mV/s; $E_i=+0.7$ V	110
6.14	Cyclic voltammogram GO_x-f -Se-MCM 41/CPE at pH 6.0 with scan rate 10 mV/s; $E_i=+700$ mV	110
6.15	Comparison of responding current of GO_x-f -MCM-41/CPE and GO_x-f -Se-MCM-41/CPE with glucose concentration using cyclic voltammetry	111
6.16	Linear voltammogram of GO_x-f - Se-MCM- 41/CPE when O_2 in graphite pores was removed during heating; $E_i=+0.7$ V	114
6.17	Linear fit of responding current and glucose concentration of GO_x-f -Se-MCM 41 with the minimized O_2 in graphite at pH 6.0 with the scan rate of 10 mV/s	116

LIST OF ABBREVIATIONS

AA	Ascorbic acid
ADA	American Diabetes Association
AOT	Sodium Bis(2-ethylhexyl) Sulfosuccinate
AP	Acetaminophen
APTES	3-aminopropyltriethoxysilane
APTMS	3-aminopropyltrimethoxysilane
<i>a</i> -Se	Amorphous Selenium
Au/GNPs-SBA-15/IO ₄ ⁻ oxidized-GOD	Gold/Gold Nanoparticles-SBA-15/ Metaperiodate Ion Oxidized- Glucose Oxidase
Au/H ₂ N-SBA-15/IO ₄ ⁻ oxidized-GOD	Gold/Amine-SBA-15/ Metaperiodate Ion Oxidized- Glucose Oxidase
Au/SWNT/GOD/PPy	Gold-Single Wall Nanotubes/ Glucose Oxidase/Polypyrrole
Au/SWNT/GOD- HRP/PPy	Gold/Single-Walled Carbon Nanotubes/Glucose Oxidase/Horseradish Peroxidase/Polypyrrole
Au/SWNT/HRP- PPy/GOD-PPy	Gold/Single Wall Nanotubes/ Horseradish Peroxidase/Polypyrrole/Glucose Oxidase/Polypyrrole
BSA	Bovine Serum albumin
CM100B	<i>Bacillus cereus</i>
CTAB	Cetyltrimethylammonium Bromide
CV	Cyclic voltammetry
d ₁₀₀	Plane 100
d ₁₁₀	Plane 110
D ₂₀₀	Plane 200
D ₂₁₀	Plane 210

DNA	Deoxyribonucleic Acid
DPCSV	Differential Pulse Cathodic Stripping Voltammetry
DTA	Differential Thermal Analysis
DTT	Dithiothreitol
E_{acc}	Accumulated potential current
EDX	Energy Dispersive X-Ray Spectroscopy
E_i	Initial potential current
Enzyme _{ox}	Enzyme oxidized
Enzyme _{red}	Enzyme reduced
<i>f</i> -Se-MCM-41	Functionalized-Selenium Nanoparticles-Mobil Crystalline Materials No. 41
Fe-MCM-41	MCM-41 Modified with Iron.
FESEM	Field Emission Scanning Electron Microscopy
<i>f</i> -MCM-41	Functionalized- Mobil Crystalline Materials No. 41
FSM-16	Folded-Sheet Mesoporous Material
FTIR	Fourier Transform Infrared Spectroscopy
G-CdS	Graphene-Cadmium Sulphur
GNPs	Gold nanoparticles
GO _x	Glucose Oxidase
GO _x - <i>f</i> -MCM-41/CPE	Glucose Oxidase-Functionalized Mobil Crystalline Materials No. 41/Carbon Paste Electrode
GO _x - <i>f</i> -MCM-41	Glucose Oxidase-Functionalized- Mobil Crystalline Materials No. 41
GO _x - <i>f</i> -Se-MCM-41	Glucose Oxidase-Functionalized Selenium Nanoparticles- Mobil Crystalline Materials No. 41
GO _x - <i>f</i> -Se-MCM-41/CPE	Glucose Oxidase-Functionalized Se-Mobil Crystalline Materials No. 41/Carbon Paste Electrode
HDP	Hydrodeporphirinization
HDTMA	Hexadecyltrimethylammonium Bromide
HMDS	Hexamethyldisilazane
ICP-MS	Inductively Coupled Plasma Mass Spectroscopy
IDDM	Insulin Dependent Diabetes Mellitus / Juvenile Diabetes
LOD	Correlative of Determination

MCF	Mesostructured Cellular Foam
MCM-41	Mobil Crystalline Materials No. 41
MCM-41-A	Amino-Mobil Crystalline Materials No. 41
MCM-41-as	As-synthesized Mobil Crystalline Materials No. 41
MCM-41-C	Calcined Mobil Mobil Crystalline Materials No. 41
MCM-41-C-1d	One day calcined Mobil Crystalline Materials No. 41
MCM-41-C-2d	Two day calcined Mobil Crystalline Materials No. 41
MCM-41-C-3d	Three day calcined Mobil Crystalline Materials No. 41
MCM-48	Mobil Crystalline Materials No. 48
MCM-50	Mobil Crystalline Materials No. 50
MOX	Malaysian Oxygen Berhad
MPTMS	3-mercaptopropyltrimethoxysilane
MWCNTs	Multi-walled carbon nanotubes
NIDDM	Non Insulin Dependent Diabetes Mellitus
PDF	Powder Diffraction File
POD	peroxidase type II from Horseradish
Pt/MCM-41	Platinum Nanoparticles/Mobil Crystalline Materials No. 41
Pt/sulfonated-MWCNTs	Platinum/sulfonated multi-walled carbon nanotubes
PTFE	Polytetraflouroethylene
PtMCWNTs	Platinum multi-walled carbon nanotubes
R^2	Correlative of determination
SBA-15	Santa Barbara Amorphous
Se-MCM-41	Selenium Nanoparticles-Mobil Crystalline Materials No. 41
SNs	Selenium Nanoparticles
β -monoclinic Se	Black crystalline Selenium (Se_8 rings)
t_{acc}	Accumulated time
TEM	Transmission Electron Microscopy
TGA	Thermal Gravimetric Analysis
TrxRs	Thioredoxinreductase
t-Se	Trigonal Selenium

UA	Uric acid
UV-Vis	Ultraviolet-visible Spectroscopy
XRD	X-ray Diffraction Spectroscopy
α -monoclinic Se	Red crystalline Selenium (Se ₈ rings)
ΔA	Absorbance changes
θ	Theta
Φ	Phi

LIST OF APPENDICES

APPENDIX	TITLE	PAGE
A1	XRD of MCM-41-as	147
A2	XRD of MCM-41-C-1d	148
A3	XRD of MCM-41-C-2d	148
A4	XRD of MCM-41-C-3d	149
A5	XRD of MCM-41-A	149
A6	XRD of <i>f</i> -MCM-41	150
B1	XRD of SNs in range 10-70°	151
B2	XRD of <i>f</i> -Se-MCM-41 in range 10-70°	152
B3	XRD of Se-MCM-41 in range 1.5-70°	152
B4	XRD of MCM-41 in range 1.5-70°	153
B5	XRD of SNs in range 1.5-70°	153
B6	XRD of Se-MCM-41 in range 1.5-70°	154
B7	XRD of <i>f</i> -Se-MCM-41 in range 1.5-70°	154
C1	FTIR of MCM-41-as	155
C2	FTIR of MCM-41-C	156
C3	FTIR of MCM-41-A	156
C4	FTIR of <i>f</i> -MCM-41	157
D1	FTIR of SNs	158
D2	FTIR of Se-MCM-41	159
D3	FTIR of <i>f</i> -Se-MCM-41	159
E1	EDX of SNs.	160
E2	EDX of Se-MCM-41	161
E3	EDX of Se-MCM-41	162

F1	Data of Se release from Se-MCM-41 within 12 h	163
F2	Percentage of Se release from Se-MCM-41 within 12 h	164
G1	Specific activity of immobilization study onto <i>f</i> -MCM-41 and <i>f</i> -Se-MCM-41 at different pH	165
G2	Percentage of specific activity of immobilization study onto <i>f</i> -MCM-41 and <i>f</i> -Se-MCM-41 at different pH	165
G3	Specific activity of immobilization study onto <i>f</i> -MCM-41 and <i>f</i> -Se-MCM-41 at different initial concentration	166
G4	Percentage of specific activity of immobilization study onto <i>f</i> -MCM-41 and <i>f</i> -Se-MCM-41 at different initial concentration	166
G5	Amount of GO _x bound onto <i>f</i> -MCM-41 and <i>f</i> -Se-MCM-41	166
G6	Percentage of GO _x bound onto <i>f</i> -MCM-41 and <i>f</i> -Se-MCM-41	167
H1	DPCSV voltammogram at pH 5.0; $t_{acc}=30$ s, $E_{acc}=+0.2$ V and $E_i=+0.7$ V. Scan rate = 50 mV/s	168
H2	DPCSV voltammogram at pH 6.0; $t_{acc}=30$ s, $E_{acc}=+0.2$ V and $E_i=+0.7$ V. Scan rate = 50 mV/s	169
H3	DPCSV voltammogram at pH 7.0; $t_{acc}=30$ s, $E_{acc}=+0.2$ V and $E_i=+0.7$ V. Scan rate = 50 mV/s	169
H4	DPCSV voltammogram at pH 9.0; $t_{acc}=30$ s, $E_{acc}=+0.2$ V and $E_i=+0.7$ V. Scan rate = 50 mV/s	170
H5	DPCSV voltammogram with scan rate 10 mV/s; pH = 6.0, $t_{acc}=30$ s, $E_{acc}=+0.2$ V and $E_i=+0.7$ V	170
H6	Linear voltammogram of GO _x - <i>f</i> -Se-MCM-41/CPE when O ₂ in graphite pores was removed during heating; $E_i=+0.7$ V	171
H7	Responding current and glucose concentration effect by pH	171
H8	Responding current and glucose concentration at	172

	scan rate of 10 mV/s	
H9	Responding current and glucose concentration at scan rate of 30 mV/s	172
H10	Responding current and glucose concentration at scan rate of 50 mV/s	172
H11	Comparison of responding current of $\text{GO}_x\text{-f-MCM-41/CPE}$ and with glucose concentration using cyclic voltammetry	173
H12	Comparison of responding current of $\text{GO}_x\text{-f-Se-MCM-41/CPE}$ with glucose concentration using cyclic voltammetry	173
H13	Responding current and glucose concentration of $\text{GO}_x\text{-f-Se-MCM 41}$ with the minimized O_2 in graphite at pH 6.0 with the scan rate of 10 mV/s	174
H14	Linear fit of responding current and glucose concentration of $\text{GO}_x\text{-f-Se-MCM 41}$ with the minimized O_2 in graphite at pH 6.0 with the scan rate of 10 mV/s	174
I1	Standard Curve of Enzyme Concentration	175
J1	Calculation of the amount of specific activity of enzyme	176

CHAPTER 1

INTRODUCTION

1.1 Background of Study

Diabetes is one of the critical diseases that are characterized by elevated glucose levels because the body cannot produce sufficient insulin or the insulin becomes resistant in regulating glucose. Worldwide, diabetes is a serious public health problem that will impact health care financing (Vinicor, 1998; Narayan *et al.*, 2000; Zhang *et al.*, 2010). Several Western countries reported that diabetic patients have increased their medical expenditure compared to individuals without diabetes from the severe macrovascular and microvascular complications associated with diabetes (Rubin *et al.*, 1994; Kangas *et al.*, 1996; Selby *et al.*, 1997; Brown *et al.*, 1999; Oliva *et al.*, 2004).

Latest data reported by the American Diabetes Association (ADA) mention that in the United States, 17 million people or 6.2% of the population were diagnosed as diabetic with 35% (5.9 millions) of these cases undiagnosed. Each year about 12,000 to 24,000 new cases of adult blindness caused by diabetes were recorded. In 1999, more than 114,000 cases of diabetes-related dialysis or transplantation that refers to end-stage renal disease were recorded. While between 1997 and 1999, 82,000 of diabetes-related amputations were recorded as non-traumatic lower extremity amputation (Hirsch, 2002).

In 1995, the number of adults with diabetes was around 135 million and this figure will rise to 300 million in the year 2025. This number, along with the

discovery of a million new cases yearly makes diabetes one of the most important national health issues that we must consider as serious and it is getting worse in developing countries because diabetes rates are increasing faster and is expected to increase 170% from 1995 to 2025 (King *et al.*, 1998). This problem lies in our lifestyle today as a result of poor nutrition intake and a sedentary lifestyle.

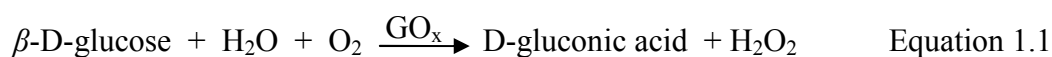
The inability of the body to control blood glucose levels can lead to acute and chronic complications. Hypoglycemia is a condition where blood glucose (glycemia) level rapidly drops to the lowest level causing mental confusion, convulsions and leading to coma or death. Chronically increased levels of glucose in the blood (chronic hyperglycemia) and an abnormally high level of proteins covalently bind with glucose (glycation or glycosylation) contributing to long-term microvascular and macrovascular complications (Lieberman and Marks, 2009).

Biosensor systems were used to detect many biological compounds especially in the field of biotechnology based on enzyme-substrate interactions. One of the applications of biosensor was to determine the levels of glucose in human body using glucose biosensor. Many researchers have developed new biosensors in order to increase sensitivity. Recently, the most popular biosensors reported are those employing redox enzyme coupled with amperometric detection because this method has the advantages of being more stable, inexpensive, simple to operate, disposable and suitable for real time detection (Chen *et al.*, 2002; Mitala Jr and Michael, 2006; Dai *et al.*, 2007; Sun *et al.*, 2007). Amperometric biosensors possess linear concentration dependence and measure changes in the current on the working electrode due to the direct oxidation of the products of a biochemical reaction in direct or indirect system. One of the key factors in the building of a reliable biosensor is the development of better techniques for immobilization of enzymes. Thus, new immobilization methods and supports are highly desired to improve the analytical capacities of sensor devices to enhance the properties such as reusability, operational stability, recovery and self life (Arai *et al.*, 2006; Jena and Raj, 2006; Park *et al.*, 2006; Shen *et al.*, 2007; Yogeswaran and Chen, 2008; Chen *et al.*, 2009; Vidotti *et al.*, 2011).

Electrochemical sensors have improved the performance of the conventional analytical tools, eliminated slow preparation, reduced the uses of expensive reagents and provided low cost analytical tools. Electrochemical sensors have certain advantages over the conventional analytical instruments such as inexpensive, portable and simple to operate. On the other hand, they also have some limitations where there are electrochemically active interferences in the sample, weak long-term stability and troublesome electron transfer pathways. However, electrochemical sensors were always applied in clinical diagnosis, environmental monitoring and food analysis.

Selenium nanoparticles (SNs) exhibit not only photoelectric, semiconductor and X-ray sensing properties, but also biological activity and good adsorptive ability due to their interaction between the SNs and N-H, C=O, COO⁻ and CN⁻ groups of the protein (Smith, and Cheatham, 1980; Ohara *et al.*, 1997; Gao *et al.*, 2002; Zhang *et al.*, 2008; Barnaby *et al.*, 2010). Hence, the SNs can be easily attached to support or enzyme and can function as a sensor that provide good amperometric signal. Many researchers have studied and synthesized stable SNs in polymer matrices (Kopeikin *et al.*, 2003a, b; Zhang *et al.*, 2007) and polysacchrides (Gao *et al.*, 2002; Zhang *et al.*, 2004a) such as chitosan (Zhang *et al.*, 2004b). MCM-41 have been found to be an exciting candidate as support for SNs compared to polymer and polysaccharides because of their uniform and adjustable pore size, defined pore and cage system, high surface area, shape and charge selectivity, high thermal stability, resistance to biodegradation and opened pore structures.

Glucose oxidase (GO_x) is a flavoprotein which catalyzes the oxidation of β-D-glucose by molecular oxygen to δ-gluconolactone, which subsequently spontaneously hydrolyzes to gluconic acid and hydrogen peroxide (Arica and Hasirci, 1993; Zoldak *et al.*, 2004).



Industrially, it is used in the removal of glucose or oxygen from food products and also applied in the production of gluconic acid (Szajani *et al.*, 1987; Ekinci *et al.*, 2007). The most important application of glucose oxidase was as a molecular diagnostic tool. The enzyme was used in biosensors for the quantitative determination of D-glucose in samples such as body fluids, foodstuffs, beverages, and fermentation products (Zoldak *et al.*, 2004). When it is applied in the voltammetry technique as a biosensor, the reaction of glucose oxidase at electrode is shown in Equation 1.3 and 1.4.



1.2 Problem Statements

Concern about health and nutrition problems is worldwide, and the detection of glucose by glucose biosensor has attracted a high degree of interest especially amongst diabetics. There are many types of glucose biosensors in the market but it is mostly too expensive. Thus, the development of a new glucose biosensor with better accuracy, high sensitivity and lower cost are needed. Today, mesoporous materials such as MCM-41, MCM-48, MCF and SBA-15 have received much attention because of their wide use in many absorbent and catalytic reactions. In addition, mesoporous materials are easy to synthesize. But mesoporous materials have their limitation in that they lack active sites for bonding with enzyme. They have to be modified to make their surface more conducive for enzyme attachment. In order to make these materials as potentially better catalysts, incorporation of metal centers such as Al, V, Fe and Mn in the silicate framework are necessary (Ozyilmaz *et al.*, 2005).

Selenium nanoparticles (SNs) was chosen not only by their unique photoelectric, semiconducting and X-ray-sensing properties but also for its biological activity and good adsorptive ability due to the interactions between the SNs and NH,

C=O, COO⁻ and C-N groups of the proteins (Smith and Cheatham, 1980; Ohara *et al.*, 1997; Gao *et al.*, 2002; Zhang *et al.*, 2008; Barnaby *et al.*, 2010). So, it can be used as a new rectifier for the component of redox enzymes based on biosensors (Zhang *et al.*, 2004b) hence chosen in the development of a more sensitive glucose biosensor

1.3 Objectives of Study

The objectives of this work are as follows:

- i. To synthesize and characterize MCM-41 and selenium nanoparticles (SNs).
- ii. To modify MCM-41 with the incorporation of SNs into MCM-41 in the presence of amino group and its characterization.
- iii. To functionalize MCM-41 and Se-MCM-41 with amino group followed by aldehyde groups and their characterization.
- iv. To optimize the immobilization of glucose oxidase enzyme (GO_x) onto functionalized MCM-41 and Se-MCM-41.
- v. To study the electrochemical properties of the GO_x-*f*-MCM-41 and GO_x-*f*-Se-MCM-41 as glucose biosensor.

1.4 Scope of Study

The scope of study covers the synthesis of MCM-41 and SNs, followed by the modification of MCM-41 with SNs in the presence of amino group (Se-MCM-41) followed by functionalization with aldehyde group (*f*-Se-MCM-41). In order to make comparison, MCM-41 was also functionalized with amino group followed by the aldehyde group. The characterization of its structural and chemical characteristics of MCM-41, SNs, Se-MCM-41 and functionalized MCM-41 (*f*-MCM-41) and Se-MCM-41 (*f*-Se-MCM-41) were investigated. The optimum conditions for the

immobilization of GO_x enzyme onto $f\text{-MCM-41}$ and $f\text{-Se-MCM-41}$ were investigated. The specific activity and percentage of GO_x bound onto $f\text{-MCM-41}$ ($\text{GO}_x\text{-}f\text{-MCM-41}$) and $f\text{-Se-MCM-41}$ ($\text{GO}_x\text{-}f\text{-Se-MCM-41}$) were determined. Then, the $\text{GO}_x\text{-}f\text{-MCM-41}$ and $\text{GO}_x\text{-}f\text{-Se-MCM-41}$ were applied as the working electrode in the electrochemical analysis (voltammetry) to determine which of these two types of electrode was suitable and feasible for the development of glucose biosensor. Particular attentions were given on the scope of electrochemical analysis which were focused to differential pulse sweep and cyclic voltammetry analysis.

1.5 Significance of Study

Glucose analysis is very important in food industry for quality control purposes, fermentation and most importantly in clinics and hospitals for diagnosing diabetic patients using glucose biosensor. This method usually involves an enzyme which is very specific for glucose and is not easily interfered by other sugars present. Due to its high selectivity towards $\beta\text{-D-glucose}$, the enzymatic method for glucose determination employs the use of glucose oxidase (GO_x). It is one of the most robust enzymes which can withstand extreme pH, ionic strength and temperature compared with other enzymes. Thus, it allows less stringent conditions during the manufacturing process and also provides relatively care-free storage which makes it more suitable for the glucose biosensors especially for home-users.

This work also deals with the modification of MCM-41 with SNs to investigate the development of a better working electrode in glucose biosensor compared to pristine MCM-41. The disadvantage of using pure mesoporous material for immobilization of enzymes is the absence of active sites in their matrices which limit the quantity of enzymes bound onto it. Besides that, incorporation of metal centers such as SNs in the silicate framework is necessary in order to increase the performance of glucose biosensor due to its semiconductor and antioxidant properties. With the incorporation of metal, Se-MCM-41 can be a suitable material for GO_x loading as well as promote electron transfer between GO_x and the electrodes.

REFERENCES

- Abdullah, A.Z., Sulaiman, N.S. and Kamaruddin, A.H. (2009). Biocatalytic Esterification of Citronellol with Lauric Acid by Immobilized Lipase on Aminopropyl-grafted Mesoporous SBA-15. *J. Biochem. Eng.* 44 , 263–270.
- Aburto, J., Ayala, M., Jaimes, I. B., Montiel, C., Terre's, E., Domínguez, J. M. and Torres, E. (2005). Stability and Catalytic Properties of Chloroperoxidase Immobilized on SBA-16 Mesoporous Materials. *Microporous and Mesoporous Mater.* 83, 193–200.
- Arai, G., Shoji, K. and Yasumori, I. (2006). Electrochemical Characteristics of Glucose Oxidase Immobilized in Poly(quinone) Redox Polymers. *J. Electroanal. Chem.* 591, 1-6.
- Araújo, R. S., Azevedo, D. C. S., Rodríguez-Catellón, E., Jiménez-López, A. and Cavalcante Jr, C. L. (2008). Al and Ti-containing Mesoporous Molecular Sieves: Synthesis, Characterization and Redox Activity in the Anthracene Oxidation. *J. Mol. Catal. A.* 281, 154-163.
- Arica, M. Y. and Bayramoğlu, G. (2004). Polyethyleneimine-Grafted Poly(hydroxyethyl methacrylate-co-glycidyl methacrylate) Membranes for Reversible Glucose Oxidase Immobilization. *J. Biochem. Eng.* 20, 73-77.
- Arica, M. Y. and Hasirci, V. (1993). Immobilization of Glucose Oxidase: A Comparison of Entrapment and Covalent Bonding. *J. Chem. Technol. Biotechnol.* 58, 287–292.

- Attia, A., Zukalova, M., Rathousky, J., Zukal, A. and Kavan, L. (2005). Mesoporous Electrode Material from Alumina-Stabilized Anatase TiO₂ for Lithium Ion Batteries. *J. Solid State Electrochem.* 9, 138–145.
- Bagnall, K.W. (1966). *The Chemistry of Selenium, Tellurium and Polonium*. Elsevier.
- Bagshaw, S.A. (2008). Rapid Calcination of High Quality Mesostructured MCM-41, MSU-X and SBA-15 Silicate Materials: A Step towards Continuous Processing? *Microporous and Mesoporous Mater.* 109, 199-209.
- Bai, Y., Yang, H., Yang, W., Li, Y. and Sun, C. (2007). Gold Nanoparticles-Mesoporous Silica Composite Used as an Enzyme Immobilization Matrix for Amperometric Glucose Biosensor Construction. *Sens. Actuators, B.* 124, 179-186.
- Barnaby, S. N., Frayne, S. H., Fath, K. R. and Banerjee, I. A. (2011). Growth of Se Nanoparticles on Kinetin Assemblies and Their Biocompatibility Studies. *Soft Mater.* 9 (4), 313–334.
- Batista, A. H. D., Sousa, F. F. D., Honorato, S. B., Ayala, A. P., Filho, J. M., Sousa, F. W. D., Pinheiro, A. N., Araujo, J. C. S. D., Nascimento, R. F., Valentini, A. and Oliveira, A. C. (2010). Ethylbenzene to Chemicals: Catalytic Conversion of Ethylbenzene into Styrene over Metal-Containing MCM-41. *J. Mol. Catal. A: Chem.* 315 (1), 86-98.
- Beck, J.S., Vartuli, J. C., Roth, W. J., Leonowicz, M. E., Kresge, C. T., Schmitt, K. D., Chu, C. T. D., Olson, D. H., Sheppard, E. W., McCullen, S. B., Higgins, J. B. and Schlenker, J. L. (1992). A New Family of Mesoporous Sieves Prepared with Liquid Crystal Templates. *J. Am. Chem. Soc.* 114, 10834-10843.
- Behrens, P. (1996). Voids in Variable Chemical Surroundings: Mesoporous Metal Oxides. *Angew. Chem. Int. Ed.* 35 (5), 515–518.

- Bhattacharyya, K. G., Talukdar, A. K., Dasa, P. and Sivasanker, S. (2003). Al-MCM-41 Catalysed Alkylation of Phenol with Methanol. *J. Mol. Catal. A:Chem.* 197, 255–262.
- Bhunia, S. and Koner, S. (2011). Functionalization of Oxo-Vanadium(IV) Acetylacetonate Over Modified MCM-41: An Efficient Reusable Catalyst for Epoxidation Reaction. *J. Porous Mater.* 18, 399–407.
- Brinchmann-Hansen, O., Dahl-Jorgensen, K., Hanssen, K.F. and Sandvik, L. (1988). The Response of Diabetic Retinopathy to 41 Months of Multiple Insulin Injections, Insulin Pumps, and Conventional Insulin Therapy. *Arch. Ophthalmol.* 106 (9), 1242–1246.
- Brown, J. B., Nichols, G. A., Glauber, H. S. and Bakst, A. W. (1999). Type 2 Diabetes: Incremental Medical Care Costs During the First 8 Years After Diagnosis. *Diabetes Care* 22, 1116-1124.
- Chanqui, C. M., Andrini, L., Fernandez, J.D., Crivello, M. E., Requejo, F. G., Herrero, E.R. and Eimer, G. A. (2010a). Speciation of Copper in Spherical Mesoporous Silicates: From the Microscale to Angstrom. *J. Phys. Chem. C* 114 (28), 12221–12229.
- Chanqui, C. M., Sapag, K., Castello, E. R., Herrero, E. R. and Eimer, G. A. (2010b). Nature and Location of Copper Nanospecies in Mesoporous Molecular Sieves. *J. Phys. Chem. C* 114 (3), 1481–1490.
- Chen, C.C., Do, J. S. and Gu, Y. (2009). Immobilization of HRP in Mesoporous Silica and its Application for the Construction of Polyaniline Modified Hydrogen Peroxide Biosensor. *Sensors* 9, 4635-4648.
- Chen, H., Shin, D. W., Nam, J. G., Kwon, K. W. and Yoo, J. B. (2010). Selenium Nanowires and Nanotubes Synthesized via a Facile Template-Free Solution Method. *Mater. Res. Bull.* 45, 699–704.

- Chen, J., Burrell, A. K., Collis, G. E., Officer, D. L., Swiegers, G. F., Too, C. O. and Wallace, G. G. (2002). Preparation, Characterisation and Biosensor Application of Conducting Polymers Based on Ferrocene Substituted Thiophene and Terthiophene. *Electrochim. Acta.* 47, 2715-2724.
- Chen, T. F., Zheng, W. J., Wong, Y. S. and Yang, F. (2008). Selenium-Induced Changes in Activities of Antioxidant Enzymes and Content of Photosynthetic Pigments in *Spirulina Platensis*. *J. Integr. Plant Biol.* 50(1), 40–48.
- Chen, Y. T., Zhang, W., Fan, Y. Q., Xu, X. Q. and Zhang, Z. X. (2006). Hydrothermal Preparation of Selenium Nanorods. *Mater. Chem. Phys.* 98, 191-194.
- Chen, Y. T., Zhang, W., Zhang, F. B., Zhang, Z. X., Zhou, B. Z. and Li, H. L. (2004). A Novel Route to Controlled Synthesis of Selenium Nanowires. *Mater. Lett.* 58, 2761-2763.
- Cheng, C. F., Zhou, W. Z., Park, D. H., Klinowski, J., Hargreaves, M. and Gladden, L. F. (1997). Controlling the Channel Diameter of the Mesoporous Molecular Sieve MCM-41. *J. Chem. Soc., Faraday Trans.* 93, 359–363.
- Chibata, I. (1978). *Immobilised Enzymes -Research and Development*. New York and London. John Wiley and Sons.
- Chong, M. A. S., Zhao, X. S., Kustedjo, A. T. and Qiao, S. Z. (2004). Functionalization of Large-Pore Mesoporous Silicas with Organosilanes by Direct Synthesis. *Microporous Mesoporous Mater.* 72, 33-42.
- Clark Jr, L. C. and Lyons, C. (1962). Electrode Systems for Continuous Monitoring in Cardiovascular Surgery. *Ann. N. Y. Acad. Sci.* 102, 29-45.
- Combs, B. S., Carper, W. R. and Stewart, J. P. (1992). The Hydrolysis of 1,5-gluconolactone-Semiempirical Methods and C-13 NMR Confirmation. *THEOCHEM.* 90 (3–4), 235–241.

- Coronado, E. and Palomares, E. (2005). Hybrid Molecular Materials for Optoelectronic Devices. *J. Mater. Chem.* 15, 3593–3597.
- Coronado, E., Galan-Mascaros, J. R., Marti-Gastaldo, C., Palomares, E., Durrant, J. R., Vilar, R., Gratzel, M. and Nazeeruddin, M. K. (2005). Reversible Colorimetric Probes for Mercury Sensing. *J. Am. Chem. Soc.* 127, 12351–12356.
- Dai, Z. H., Ni, J., Huang, X. H., Lu, G. F. and Bao, J. C. (2007). Direct Electrochemistry of Glucose Oxidase Immobilized on a Hexagonal Mesoporous Silica-MCM-41 Matrix. *Bioelectrochem.* 70 (2), 250-256.
- Deera, J., Magner, E., Wall, J. G. and Hodnett, B. K. (2002). Mechanistic and Structural Features of Protein Adsorption onto Mesoporous Silicates. *J. Phys. Chem. B.* 106, 7340-7347.
- DeWitt, D. E. and Hirsch, I. B. (2003). Outpatient Insulin Therapy in Type 1 and Type 2 Diabetes Mellitus: Scientific Review. *JAMA.* 289 (17), 2254-2264.
- Dhanjal, S. and Cameotra, S. S. (2010). Aerobic Biogenesis of Selenium Nanospheres by *Bacillus cereus* Isolated from Coalmine Soil. *Microb. Cell Fact.* 9 (1), 52.
- Diaz, J. F. and Balkus Jr, K. J. (1996). Enzyme Immobilization in MCM-41 Molecular Sieve. *J. Mol. Catal. B.* 2, 115-126.
- Eggins, B. R. (1996). *Biosensors: An Introduction*. England: John Wiley & Sons Ltd.
- Ekinci, O., Boyukbayram, E., Kiralp, S., Toppare, L. and Yagci, Y. (2007). Characterization and Potential Applications of Immobilized Glucose Oxidase and Polyphenol Oxidase. *J. Macromol. Sci. Part A Pure Appl. Chem.* 44, 801–808.

- Eriksson, K. O., Kourteva, I., Yao, K., Liao, J. L., Kilar, F. and Hjerten, S. (1987). Application of High-Performance Chromatographic and Electrophoretic Methods to the Purification and Characterization of Glucose Oxidase and Catalase from *Penicillium Chrysogenum*. *J. Chromatogr.* 397, 239–49.
- Gao, X., Zhang, J. and Zhang, L. (2002). Hollow Sphere Selenium Nanoparticles. Their In-Vitro Anti-Hydroxyl Radical Effect. *Adv. Mater.* 14, 290-293.
- Gates B. and Xia, Y. N. (2000). A Solution-Phase Approach to the Synthesis of Uniform Nanowires of Crystalline Selenium with Lateral Dimensions in the Range of 10-30 nm. *J. Am. Chem. Soc.* 122, 12582-12583.
- Gates, B., Mayers, B., Cattle, B. and Xia, Y. (2002). Synthesis and Characterization of Uniform Nanowires of Trigonal Selenium. *Adv. Func. Mater.* 12, 219-223.
- Gautam, U. K., Nath, M. and Rao, C. N. R. (2003). New Strategies for the Synthesis of t-Selenium Nanorods and Nanowires. *J. Mater. Chem.* 13, 2845-2847.
- Gratzel, M. (2005). Solar Energy Conversion by Dye-Sensitized Photovoltaic Cells. *Inorg. Chem.* 44, 6841–6851.
- Guan, L. C. (2005). *Mesoporous MCM-48 Synthesized from Rice Husk Ash Silica: Physicochemical Properties and its Catalytic Activity in Acylation Reaction*. MSc. Thesis. Universiti Teknologi Malaysia, Skudai.
- Guidini, C. Z., Fischer, J., Santana, L.N.S., Cardoso, V.L. and Ribeiro, E.J. (2010). Immobilization of *Aspergillus Oryzae* β -Galactosidase in Ion Exchange Resins by Combined Ionic-Binding Method and Cross-linking. *J. Biochem. Eng.* 52, 137–143.
- Hartmann, M. (2005). Ordered Mesoporous Materials for Bioadsorption and Biocatalysis. *Chem. Mater.* 17, 4577-4593.

- He, J., Yang, X., Evans, D. G. and Duan, X. (2001). New Methods to Remove Organic Templates from Porous Materials. *Mater. Chem. Phys.* 1 (77), 270-275.
- Heilman, A., Teuscher, N., Kiesow, A., Janasek, D. and Spohn, U. (2003). Nanoporous Aluminium Oxide as a Novel Support Material for Enzyme Biosensor. *J. Nanosci. Nanotechnol.* 3, 357-379.
- Heller, S. R., Colagiuri, S., Vaaler, S., Wolffenbuttel, B. H., Koelendorf, K., Friberg, H. H., Windfeld, K. and Lindholm, A. (2004). Hypoglycaemia with Insulin Aspart: A Double-Blind, Randomised, Crossover Trial in Subjects with Type 1 Diabetes. *Diabet. Med.* 21 (7), 769-775.
- Henning, T. P. and Cunningham, D. D. (1998). *Commercial Biosensors: Applications to Clinical Bioprocess and Environmental Samples*. New York. John Wiley & Sons.
- Hirsch, I. B. (2002). The Prevention of Type 2 Diabetes: Are We Ready for the Challenge?. *Clin. Diabetes* 20, 106-108.
- Hodak, J., Etchenique, R., Calvo, E. J., Singhal, K. and Bartlett, P. N. (1997). Layer-By Layer Self-Assembly of Glucose Oxidase with a Poly(allylamine)ferrocene Redox Mediator. *Langmuir*. 13, 2708–2716.
- Hudson, S., Magner, E., Cooney, J. and Hodnett, B. K. (2005). Methodology for the Immobilization of Enzymes onto Mesoporous Materials. *J. Phys. Chem. B.* 109, 19496-19506.
- Ispas, C., Sokolov, I. and Andreescu, S. (2009). Enzyme-Functionalized Mesoporous Silica for Bioanalytical Applications. *Anal. Bioanal. Chem.* 393, 543-554.
- Jena, B. K. and Raj, C. R. (2006). Enzyme-Free Amperometric Sensing of Glucose by Using Gold Nanoparticles. *Chem. Eur. J.* 12, 2702-2708.

- Jung, D. and Hartmann, M. (2010). Oxidation of Indole with CPO and GOx Immobilized on Mesoporous Molecular Sieves. *Catal. Today* 157, 378–383.
- Jung, D., Streb, C. and Hartmann, M. (2010). Covalent Anchoring of Chloroperoxidase and Glucose Oxidase on the Mesoporous Molecular Sieve SBA-15. *Int. J. Mol. Sci.* 11, 762-778.
- Kalisz, H. M., Hecht, H. J., Schomburg, D. and Schmid, R. D. (1991). Effects of Carbohydrate Depletion on the Structure, Stability and Activity of Glucose Oxidase from *Aspergillus Niger*. *Biochim. Biophys. Acta.* 1080 (2), 138–142.
- Kangas, T., Aro, S., Koivisto, V. A., Salinto, M., Laakso, M. and Reunanen, A. (1996). Structure and Costs of Health Care of Diabetic Patients in Finland. *Diabetes Care* 19, 494–497.
- Kim, J., Jia, H. and Wang, P. (2006). Challenges in Biocatalysis for Enzyme-Based Biofuel Cells. *Biotechnol. Adv.* 24, 296– 308.
- King, H., Aubert, R. E. and Herman, W. H. (1998). Global Burden of Diabetes, 1995-2025: Prevalence, Numerical Estimates and Projections. *Diabetes Care.* 21 (9), 1414-1431.
- Kinsel, M. E. G., Jimenez, V. L., Washmon, L. and Balkus Jr, K. J. (1998). Mesoporous Molecular Sieve Immobilized Enzymes. *Stud. Surf. Sci. Catal.* 117, 373-380.
- Koide, S. and Yokoyama, K. (1999). Electrochemical Characterization of an Enzyme Electrode Based on a Ferrocene Containing Redox Polymer. *J. Electroanal. Chem.* 468, 193-201.
- Kopeikin, V. V., Valueva S. V., Kipper A. I., Borovikova, L. N., Nazarkina, Y. I. Khlebosolova, E. N. and Filippov, A. I. (2003a). Adsorption of Hydroxyethyl Cellulose Selenium Nanoparticles During Their Formation in Water. *Russ. J. Appl. Chem.* 76 (4), 600-602.

- Kopeikin, V. V., Valueva, S. V., Kipper, A. I., Borovikova, L. N., and Filippov, A. I. (2003b). Synthesis of Selenium Nanoparticles in Aqueous Solutions of Poly(vinylpyrrolidone) and Morphological Characteristics of The Related Nanocomposites. *Polym. Sci. A*. 45, 374-379.
- Kresge, C. T., Leonowicz, M. E., Roth, W. J., Vartuli, J. C. and Beck, J. S. (1992). Ordered Mesoporous Molecular Sieves Synthesized by a Liquid-Crystal Template Mechanism. *Nature*. 359 (6937), 710-712.
- Kriel, L. W., Jimenez, V. L. and Balkus Jr, K. J. (2000). Cytochrome *c* Immobilization into Mesoporous Molecular Sieves. *J. Mol. Catal. B: Enzym.* 10, 453–469.
- Langi, B., Shah, C., Singh, K., Chaskar, A., Kumar, M. and Bajaj, P. N. (2010). Ionic Liquid-Induced Synthesis of Selenium Nanoparticles. *Mater. Res. Bull.* 45 (6), 668-671.
- Lee, C. H., Lin, T. S. and Mou, C. Y. (2009). Mesoporous Materials for Encapsulating Enzymes. *Nano. Today* 4, 165—179.
- Lee, H. S., Kim, W. H., Lee, J. H., Choi, D. J., Jeong, Y. K. and Chang, J. H. (2012). Transition Metal-Chelating Surfactant Micelle Templates for Facile Synthesis of Mesoporous Silica Nanoparticles. *J. Solid State Chem.* 185, 89-94.
- Lei, C., Shin, Y., Liu, J. and Ackerman, E. J. (2002). Entrapping Enzyme in a Functionalized Nanoporous Support. *J. Am. Chem. Soc.* 124, 11242-11243.
- Lemus, I. A., Gómez, Y. V. and Contreras, L. A. (2011). Platinum Nanoparticles Synthesis Supported in Mesoporous Silica and its Effect in MCM-41 Lattice. *Int. J. Electrochem. Sci.* 6, 4176 – 4187.
- Li, X. M., Li, Y., Li, S. Q., Zhou, W. W., Chu, H. B., Chen, W., Li, I. L. and Tang, Z. (2005). Single Crystalline Trigonal Selenium Nanotubes and Nanowires Synthesized by Sonochemical Process. *Cryst. Growth Des.* 3 (5), 911-916.

- Lieberman, M. and Marks, A. D. (2009). *Marks' Basic Medical Biochemistry: A Clinical Approach Third Edition*. Philadelphia. Lippincott Williams & Wilkins.
- Lin, K. C., Tsai, T. H. and Chen, S. M. (2010). Performing Enzyme-Free H_2O_2 Biosensor and Simultaneous Determination for AA, DA, and UA by MWCNT–PEDOT Film. *Biosens. and Bioelectron.* 26, 608–614.
- Linares, C. F., Amézqueta, P. and Scott, C. (2008). Mo/MCM-41-Type Mesoporous Materials Doubly Promoted with Fe and Ni for Hydrotreating Reactions. *Fuel* 87 (12), 2817–2823.
- Liu, B. S., Xu, D. F., Chu, J., X., Liu, W. and Au, C. T. (2007). Deep Desulfurization by the Adsorption Process of Fluidized Catalytic Cracking (FCC) Diesel over Mesoporous Al–MCM-41 Materials. *Energy Fuels* 21 (1), 250-255.
- Liu, B., Cao, Y., Chen, D., Kong, J. and Deng, J. (2003). Amperometric Biosensor Based on a Nanoporous ZrO_2 Matrix. *Anal. Chim Acta.* 478, 59-66.
- Liu, B., Hu, R. and Deng, J. (1997). Characterization of Immobilization of an Enzyme in a Modified Y Zeolite Matrix and its Application to an Amperometric Glucose Biosensor. *Anal. Chem.* 69, 2343-2348.
- Liu, L., Peng, Q. and Li, Y. (2008). Preparation of Monodisperse Se Colloid Spheres and Se Nanowires Using Na_2SeSO_3 as Precursor. *Nano. Res.* 1, 403-411.
- Liu, M. Z., Zhang, S. Y., Shen, Y. H. and Zhang, M. L. (2004). Selenium Nanoparticles Prepared from Reverse Microemulsion Process. *Chin. Chem. Lett.* 15 (10), 1249-1252.
- Lu, J., Xie, Y., Xu, F., and Zhu, L. (2002). Study of the Dissolution Behavior of Selenium and Tellurium in Different Solvents - A Novel Route to Se, Te Tubular Bulk Single Crystal. *J. Mater. Chem.* 12, 2755-2761.

- Mateo, C., Palomo, J. M., Fernandez-Lorente, G., Guisan, J. M. and Fernandez-Lafuente, R. (2007). Improvement of Enzyme Activity, Stability and Selectivity via Immobilization Techniques. *Enzyme Microb. Technol.* 40 (6), 1451–63.
- Mehta, S. K., Chaudhary, S., Kumar, S., Bhasin, K. K., Torigoe, K., Sakai, H. and Masahiko, A. (2008). Surfactant Assisted Synthesis and Spectroscopic Characterization of Selenium Nanoparticles in Ambient Conditions. *Nanotechnology* 19 (29), 295601.
- Melde, B. J., Johnson, B. J. and Charles, P. T. (2008). Mesoporous Silicate Materials in Sensing. *Sensors*. 8, 5202-5228.
- Mitala Jr, J. J., and Michael, A. C. (2006). Improving the Performance of Electrochemical Microsensors Based on Enzymes Entrapped in a Redox Hydrogel. *Anal. Chim. Acta.* 556, 326-332.
- Mohamed, A. (2005) *Synthesis, Characterization and Activity of Al-MCM-41 Catalyst for Hydroxyalkylation of Epoxides*. MSc. Thesis. Universiti Teknologi Malaysia, Skudai.
- Montiel, C., Terr'es, E., Dom'nguez, J. and Aburto, J. (2007). Immobilization of Chloroperoxidase on Silica-Based Materials for 4,6-dimethyl dibenzothiophene Oxidation. *J. Mol. Catal. B: Enzym.* 48, 90–98.
- Mukti, R. R. (2003). *H-Al-MCM-41 in the Benzoylation of Biphenyl for the Formation of Disubstituted 4, 4'-dibenzoylbiphenyl*. MSc.Thesis. Universiti Teknologi Malaysia, Skudai.
- Mullie F. L., Pauw, D. D. and Lepoint, T. (1996). Analysis of the 'New Electrical Model' of Sonoluminescence. *Ultrason. Sonochem.* 3, 73-76.
- Newman, J. D. and Turner, A. P. F. (2005). Home Blood Glucose Biosensors: A Commercial Perspective. *Biosens. Bioelectron.* 20, 2435-2453.

- Niu, J. and Lee, J. L. (2002). Reagentless Mediated Biosensors Based on Polyelectrolyte and Sol-gel Derived Silica Matrix. *Sens. Actuators, B.* 82, 250-258.
- Norouzian, D. (2003). Enzyme Immobilization: The State of Art in Biotechnology. *Iran. J. Biotechn.* 4 (1), 197-206.
- Ohara, P., Gelbart., W. M. and Heath, J. R. (1997). Self-Assembly of Sub-Micrometer Rings of Particles from Solutions of Nanoparticles. *Ang. Chem. Comm. Int. Ed. Engl.* 36, 1078-1080.
- Ohinmaa, A., Jacobs, P., Simpson, S. and Johnson, J. A. (2004). The Projection of Prevalence and Cost of Diabetes in Canada: 2000 to 2016. *Can. J. Diabetes.* 28, 116-123.
- Ohkubo, Y., Kishikawa, H., Araki, E., Miyata, T., Isami, S., Motoyoshi, S., Kojima, Y., Furuyoshi, N. and Shichiri, M. (1995). Intensive Insulin Therapy Prevents the Progression of Diabetic Microvascular Complications in Japanese Patients with Non-insulin-dependent Diabetes Mellitus: A Randomized Prospective 6-years Study. *Diabetes Res. Clin. Pract.* 28 (2), 103-117.
- Oliva, J., Lobo, F., Molina, B. and Monereo, S. (2004). Direct Health Care Costs of Diabetic Patients in Spain. *Diabetes Care* 27, 2616-2621.
- Ozyilmaz, G., Tukul, S. S. and Alptekin, O. (2005). Activity and Storage Stability of Immobilized Glucose Oxidase onto Magnesium Silicate. *J. Mol. Catal. B: Enzym.* 35, 154-160.
- Pandya P. H., Jasra R. V., Newalkar B. L. and Bhatt P. N. (2005). Studies on the Activity and Stability of Immobilized α -Amylase in Ordered Mesoporous Silicas. *Microporous Mesoporous Mater.* 77, 67-77.
- Park, S., Boo, H. and Chung, T. D. (2006). Electrochemical Non-Enzymatic Glucose Sensors. *Anal. Chim. Acta.* 556, 46-57.

- Perenlei, G., Tee, T. W., Yusof, N. A. and Kheng, G. J. (2010). Voltammetric Detection of Potassium Ferricyanide Mediated by Multi-walled Carbon Nanotube/Titanium Dioxide Composite Modified Glassy Carbon Electrode. *Int. J. Electrochem. Sci.* 6, 520 – 531.
- Pérez, L. D., López, J. F, Orozco, V. H., Kyu, T. and López, B. L. (2008). Effect of the Chemical Characteristics of Mesoporous Silica MCM-41 on Morphological, Thermal and Rheological Properties of Composites Based on Polystyrene. *J. Appl. Polym. Sci.* 111, 2229-2237.
- Petri, A., Gambicorti, T. and Salvadori, P. (2004). Covalent Immobilization of Chloroperoxidase on Silica Gel and Properties of the Immobilized Biocatalyst. *J. Mol. Catal. B: Enzym.* 27, 103-106.
- Plank, J., Siebenhofer, A., Berghold, A., Jeitler, K., Horvath, K., Mrak, P. and Pieber, T. R. (2005). Systematic Review and Meta-Analysis of Short-Acting Insulin Analogues in Patients with Diabetes Mellitus. *Arch. Intern. Med.* 165, 1337-1344.
- Popova, M., Szegedi, Á. Lázár, K. and Dimitrova, A. (2011). Dehydrogenation of Cyclohexanol on Fe, Ti-MCM-41 Mesoporous Materials. *Catal. Lett.* 141, 1288–1296.
- Prado, A. G. and Airoidi, C. (2001). Adsorption, Preconcentration and Separation of Cations on Silica Gel Chemically Modified with the Herbicide 2,4-Dichlorophenoxyacetic Acid. *Anal. Chim. Acta.* 432(2), 201-211.
- Rauf, S., Ihsan, A., Akhtar, K., Ghauri, M. A., Rahman, M., Anwar, M. A. and Khalid, A. M. (2006). Glucose Oxidase Immobilization on a Novel Cellulose Acetate-Polymethylmethacrylate Membrane. *J. Biotechnol.* 121, 351-360.
- Reichard, P., Nilsson, B. Y. and Rosenqvist, U. (1993). The Effect of Long-Term Intensified Insulin Treatment on the Development of Microvascular Complications of Diabetes Mellitus. *N. Engl. J. Med.* 329, 304-309.

- Reynolds, E. R., Geise, J. R. and Yacynych, A. M. (1992). *Biosensor & Chemical Sensors : Optimization Performance Through Polymeric Material*. Edelman, P. G and Joseph Wang. USA: Library of Congress.
- Rossi, L. M., Quach, A. D. and Rosenzweig, Z. (2004). Glucose Oxidase/magnetite Nanoparticle Bioconjugate for Glucose Sensing. *Anal. Bioanal. Chem.*, 380 (4), 606.
- Rubin R. J., Altman, W. M. and Mendelson, D. N. (1994). Health Care Expenditures for People with Diabetes Mellitus. *J. Clin. Endocrinol. Metab.* 78, 809A-809F.
- Samadi-Maybodi, A., Teymouri, M., Vahid, A. and Miranbeigi, A. (2011). In Situ Incorporation of Nickel Nanoparticles into the Mesopores of MCM-41 by Manipulation of Solvent-Solute Interaction and its Activity toward Adsorptive Desulfurization of Gas Oil. *J. Hazard Mater.* 192 (3), 1667-1674.
- Sanjay, G. and Sugunan, S. (2006). Enhanced pH and Thermal Stabilities of Invertase Immobilized on Montmorillonite K-10. *Food Chem.* 94, 573–579.
- Selby, J. V., Ray, G. T., Zhang, D. and Colby, C. J. (1997). Excess Costs of Medical Care for Patients with Diabetes in a Managed Care Population. *Diabetes Care* 20, 1396-1402.
- Shen, J., Dudik, L. and Liu, C. C. (2007). An Iridium Nanoparticles Dispersed Carbon Based Thick Film Electrochemical Biosensor and its Application for a Single Use, Disposable Glucose Biosensor. *Sens. Actuat. B.* 125, 106-113.
- Siebenhofer, A., Plank, J., Berghold, A., Jeitler, K., Horvath, K., Narath, M., Gfrerer, R., and Pieber, T. R. (2006). Short Acting Insulin Analogues versus Regular Human Insulin in Patients with Diabetes Mellitus. CD003287. *Cochrane Database Syst. Rev.* 2, 4. University of Medicine, Austria.
- Sinaga, S. M. (2002). Penggunaan Voltammetri Elektrod Pasta Karbon dan Elektrod Pasta Karbon Terubahsuai Bagi Penentuan Sebatian Asid 3-Nitro-4-

Hidrosifenilarsonik dan Asid para Arsanilik. PhD Thesis. Universiti Teknologi Malaysia, Skudai.

Siriluk, C. and Yuttapong, S. (2005). Structure of Mesoporous MCM-41 Prepared from Rice Husk Ash. *8TH Asian Symposium on Visualization*. 23-27 May 2005. Chiangmai, Thailand.

Sirvent, M. A., Merkoci, A. and Alegret, S. (2000). Configurations Used in the Design of Screen-Printed Enzymatic Biosensors: A Review. *Sens. Actuators, B*. 69, 153-163.

Slowing, I.I., Trewyn, B.G., Giri, S. and Lin, V.S.Y. (2007). Mesoporous Silica Nanoparticles for Drug Delivery and Biosensing Applications. *Adv. Funct. Mater.* 17, 1225-1236.

Smith, T. W. and Cheatham, R. A. (1980). Functional Polymers in the Generation of Colloidal Dispersions of Amorphous Selenium. *Macromolecules* 13 (5), 1203-1207.

Souness, J. E. and Stouffer, J. E. (1983). The Effect of Selenium-Deficiency on Rat Fat-cell Glucose Oxidation. *Biochem. J.* 214, 471-477.

Stefanis, A. D., Kaciulis, S. and Pandolfi, L. (2007). Preparation and Characterization of Fe-MCM-41 Catalysts Employed in the Degradation of Plastic Materials. *Microporous Mesoporous Mater.* 99, 140-148.

Straumal, B. B., Gust, W., Molodov, D. A. (1995). Wetting Transition on Grain Boundaries in Al Contacting with a Sn-Rich Melt. *Interface Sci.* 3, 127-123.

Suhail, M. and Rizvi, S. I. (1989). Erythrocyte Membrane Acetylcholinesterase in Type 1 (Insulin-Dependent) Diabetes Mellitus. *Biochem. J.* 259, 897-899.

Sun, Y., Bai, Y., Yang, W. and Sun, C. (2007). Controlled Multilayer Films of Sulfonate-Capped Gold Nanoparticles/Thionine Used for Construction of a

- Reagentless Bienzymatic Glucose Biosensor. *Electrochim. Acta.* 52, 7352-7361.
- Suslick, K. S. (1990). Sonochemistry. *Science.* 247, 1439-1445.
- Suslick, K. S., Doktycz, S. J. and Flint., E. B. (1990). On the Origin of Sonoluminescence and Sonochemistry. *Ultrasonics.* 5 (28), 280-290.
- Szajani, B., Molnar, A., Klamar, B. and Kalman, M. (1987). Preparation, Characterization and Potential Application of an Immobilized Glucose Oxidase. *Appl. Biochem. Biotechnol.* 14, 37-47.
- Takahashi, H., Li B., Sasaki, T., Miyazaki, C., Kajino, T. and Inagaki, S. (2000). Catalytic Activity in Organic Solvents and Stability of Immobilized Enzymes Depend on the Pore Size and Surface Characteristics of Mesoporous Silica. *Chem. Mater.* 12, 3301-3305.
- Takahashi, H., Li, B., Sasaki, T., Miyazaki, C., Kajino, T. and Inagaki, S. (2001). Immobilized Enzymes in Ordered Mesoporous Silica Materials and Improvement of their Stability and Catalytic Activity in an Organic Solvent. *Microporous and Mesoporous Mater.* 44-45, 755-762.
- Tanne, J., Schäfer, D., Khalid, W., Parak, W.J. and Lisdat, F. (2011). Light-Controlled Bioelectrochemical Sensor Based on CdSe/ZnS Quantum Dots. *Anal. Chem.* 83 (20), 7778-7785.
- Tchobroutsky, G. (1978). Relation of Diabetic Control to the Development of Microvascular Complications. *Diabetologia* 15, 143-152.
- The Diabetes Control and Complications Trial Research Group. (1993). The Effect of Intensive Treatment of Diabetes on the Development and Progression of Long-Term Complications in Insulin-Dependent Diabetes Mellitus. *N. Engl. J. Med.* 329, 977-986.

- Tian, Z. R., Tong, W., Wang, J. Y., Duan, N. G., Krishnan, V. V. and Suib, S. L. (1997). Manganese Oxide Mesoporous Structures: Mixed-Valent Semiconducting Catalysts. *Science*. 276 (5314), 926-930.
- Trewyn, B.G., Giri, S., Slowing, I. I, and Lin, V. S. Y. (2007). Mesoporous Silica Nanoparticle Based Controlled Release, Drug Delivery and Biosensor System. *Chem. Commun.*, 3236-3245.
- Tsoncheva, T., Järn, M., Paneva, D., Dimitrov, M. and Mitov, I. (2011). Copper and Chromium Oxide Nanocomposite Supported on SBA-15 Silica as Catalyst for Ethylacetate Combustion: Effect of Mesoporous Structure and Metal Oxide Composition. *Microporous Mesoporous Mater.* 137 (1-3), 56-64.
- Urdike, S. J. and Hicks, G. P. (1967a). The Enzyme Electrode. *Nature*. 214, 986-988.
- Urdike, S. J. and Hicks, G. P. (1967b). Reagentless Substrate Analysis with Immobilized Enzymes. *Science* 158, 270-272.
- Vartuli, J. C., Schmitt, K. D., Kresge, C. T., Roth, W. J., Leonowicz, M. E., McCullen, S. B. Hellring, S. D., Beck, J. S., Schlenker, J. L., Olson, D. H. and Sheppard, E. W. (1994). Effect of Surfactant Silica Molar Ratios on the Formation of Mesoporous Molecular-Sieves—Inorganic Mimicry of Surfactant Liquid-Crystal Phases and Mechanistic Implications. *Chem. Mater.* 6, 2317–2326.
- Velu, S., Wang, L., Okazaki, M., Suzuki, K. and Tomura, S. (2002). Characterization of MCM-41 Mesoporous Molecular Sieves Containing Copper and Zinc and their Catalytic Performance in the Selective Oxidation of Alcohols to Aldehydes. *Microporous Mesoporous. Mater.* 54 (1–2), 113–126.
- Vidotti, M., Carvalhal, R. F., Mendes, R. K., Ferreira, D. C. M. and Kubota, L. T. (2011). Biosensors Based on Gold Nanostructures. *J. Braz. Chem. Soc.* 22 (1), 3-20.

- Vinikor, F. (1998). The Public Health Burden of Diabetes and the Reality of Limits. *Diabetes Care* 21 (3), 15-8.
- Vinu, A., Murugesan, V., Hartmann, M. (2004a). Adsorption of Lysozyme over Mesoporous Molecular Sieves MCM-41 and SBA-15: Influence of pH and Aluminium Incorporation. *J. Phy. Chem B.* 108, 7323-7330.
- Vinu, A., Murugesan, V., Tangermann, O., Hartmann, M. (2004b). Adsorption of Cytochrome c on Mesoporous Molecular Sieves: Influence of pH, Pore Diameter and Aluminium Incorporation. *Chem Mater.* 16, 3056-3065.
- Walcarius, A. (2008). Electroanalytical Applications of Microporous Zeolites and Mesoporous (Organo)silicas: Recent Trends. *Electroanalysis.* 20, 711-738.
- Walcarius, A., Mandler, D., Cox, J. A., Collinson, M. and Lev, O. (2005). Exciting New Directions in the Intersection of Functionalized Sol–Gel Materials with Electrochemistry. *J. Mater. Chem.* 15, 3663–3689.
- Wang, H. J., Zhou, C. M., Peng, F. and Yu, H. (2007). Glucose Biosensor Based on Platinum Nanoparticles Supported Sulfonated-Carbon Nanotubes Modified Glassy Carbon Electrode. *Int. J. Electrochem. Sci.* (2), 508-516.
- Wang, K., Liu, Q., Guan, Q. M., Wu, J., Li, H. N. and Yan, J. J.(2011). Enhanced Direct Electrochemistry of Glucose Oxidase and Biosensing for Glucose via Synergy Effect of Graphene and CdS Nanocrystals. *Biosens. Bioelectron.* 26 (5), 2252-2257.
- Wang, X., Zheng, X., Lu, J. and Xie, Y. (2003). Reduction of Selenious Acid Induced by Ultrasonic Irradiation–Formation of Se Nanorods. *Ultrason. Sonochem.* 11 (5), 307-310.
- Wang, Y. and Caruso, F. (2004). The MBs are Expected to be Immobilized in the 10-40 nm Pores. *Chem. Commun.* 1528-1529.

- Wang, Y. and Caruso, F. (2005). Mesoporous Silica Spheres as Supports for Enzyme Immobilization and Encapsulation. *Chem. Mater.* 17, 953-961.
- Wang, Y., Xu, H., Zhang, J. and Li, G. (2008). Electrochemical Sensors for Clinic Analysis. *Sensors* 8, 2043-2081.
- Wei, Y., Xu, J., Feng, Q., Dong, H. and Lin, M. (2000). Encapsulation of Enzymes in Mesoporous Host Materials via the Nonsurfactant-templated Sol-gel Process. *Mater.Lett.* 44(1), 6-11.
- Wei, Y., Xu, J., Feng, Q., Lin, M., Dong, H., Zhang, W., Wang, C. (2001). A Novel Method for Enzyme Immobilization: Direct Encapsulation of Acid Phosphatase in Nanoporous Silica Host Materials. *J. Nanosci. Nanotechnol.* 1(1), 83-93.
- Williams, C. D. (2009). *Personal communication*. University of Wolverhampton, United Kingdom.
- Xi, G., Xiong, K., Zhao, Q., Zhang, R., Zhnag, H. and Qian, Y. (2006). Nucleation-Dissolution-Recrystallization: A New Growth Mechanism for *t*-Selenium Nanotubes. *Crys.Growth Des.* 6 (2), 577-582.
- Xie, T., Wang, A., Huang, L., Li, H., Chen, Z., Wang, Q. and Yin, X. (2009). Recent Advance in the Support and Technology Used in Enzyme Immobilization. *Afr. J. Biotech.* 8 (19), 4724-4733.
- Yang, Y. M., Wang, J. W. and Tan, R. X. (2004). Immobilization of Glucose Oxidase on Chitosan-SiO₂ gel. *Enzyme. Microb. Technol.* 34, 126-131.
- Yiu, H. H. P., Wright, P. A. and Botting, N. P. (2001a). Enzyme Immobilisation using Siliceous Mesoporous Molecular Sieves. *Microporous Mesoporous Mater.* 44-45, 763-768.

- Yiu, H. H. P., Wright, P. A. and Botting, N. P. (2001b). Enzyme Immobilisation using SBA-15 Mesoporous Molecular Sieves with Functionalised Surfaces. *Journal of Molecular Catalysis B*. 15, 81–92.
- Yogeswaran, U. and Chen, S. M. (2008). A Review on the Electrochemical Sensors and Biosensors Composed of Nanowires as Sensing Material. *Sensors* 8, 290-313.
- Yoon, H. C. and Kim, H. S. (2000). Multilayered Assembly of Dendrimers with Enzymes on Gold: Thickness-Controlled Biosensing Interface. *Anal. Chem.* 72, 922-826.
- Yoon, H. C., Hong, M. Y. and Kim, H.S. (2000). Functionalization of a Polyamidoamine Dendrimer with Ferrocenyls and its Application to the Construction of a Reagentless Enzyme Electrode. *Anal. Chem.* 72, 4420-4427.
- Zhang, J., Zhang, S. Y., Xu, J. J. and Chen, H. Y. (2004a). A New Method for the Synthesis of Selenium Nanoparticles and the Application to Construction of H₂O₂ Biosensor. *Chin. Chem. Lett.* 15 (11), 1345-1348.
- Zhang, S. Y., Zhang, J., Wang, H. Y. and Chen, H. Y. (2004b). Synthesis of Selenium Nanoparticles in the Presence of Polysaccharides. *Mater. Lett.* 58 (21), 2590-2594.
- Zhang, B., Hou, W., Zhu, X., Ye, X., Fei, L., Liu, X., Yang, J. and Xie, Y. (2007). Multiarmed Tubular Selenium with Potentially Unique Electrical Properties: Solution-Phase Synthesis and First- Principles Calculation. *Small* 3 (1), 101-105.
- Zhang, J., Wang, X. and Xu, T. (2008). Elemental Selenium at Nano Size (Nano-Se) as a Potential Chemopreventive Agent with Reduced Risk of Selenium Toxicity: Comparison with Se-Methylselenocysteine in Mice. *Toxicol. Sci.* 101(1), 22–31.

- Zhang, J., Lin, L. Zhang, J. and Shi, J. (2011). Efficient Conversion of D-glucose into D-Sorbitol over MCM-41 Supported Ru Catalyst Prepared by a Formaldehyde Reduction Process. *Carbohydr. Res.* 346 (11), 1327-1332.
- Zhao, W., Xu, J. J., Qiu, Q. Q. and Chen, H. Y. (2006). Nanocrystalline Diamond Modified Gold Electrode for Glucose Biosensing. *Biosens. Bioelectron.* 22, 649-655.
- Zhou, J. Q. (2010). Immobilization of Cellulase on a Reversibly Soluble-Insoluble Support: Properties and Application. *J. Agric. Food Chem.* 58, 6741–6746.
- Zhu, L., Yang, R., Zhai, J. and Tian, C. (2007). Bienzymatics Glucose Biosensor Based on Co-Immobilization of Peroxidase and Glucose Oxidase on a Carbon Nanotubes Electrode. *Biosens. Bioelectron.* (23), 528-535.
- Zhu, W., Xu, H., Wang, W. and Shi, J. (2006). Controlled Synthesis of Trigonal Selenium Nanowires via a Facile Solution Route. *Appl. Phys. A.* 83, 281-284.
- Zingaro, R. A. and Cooper, W.C. (1974). *Selenium*. New York/ Cincinnati/ Toronto/ London/ Melbourne: Van Nostrand Reinhold Company.
- Zoldak, G., Zubrik, A., Musatov, A., Stupak, M. and Sedlak, E. (2004). Irreversible Thermal Denaturation of Glucose Oxidase from *Aspergillus Niger* is the Transition to the Denatured State with Residual Structure. *J. Biol. Chem.* 279, 47601-47609.